## A TCR-pMHC confinement time model of T cell activation

Daniel Coombs

Department of Mathematics & Institute of Applied Mathematics

University of British Columbia

#### Acknowledgements

#### **University of British Columbia:**

Omer Dushek

Raibatak Das

#### **Oxford University:**

Omer Dushek

Anton van der Merwe

Milos Aleksic

#### Los Alamos:

Byron Goldstein

#### Chile:

Pablo Gonzalez

Alexis Kalergis



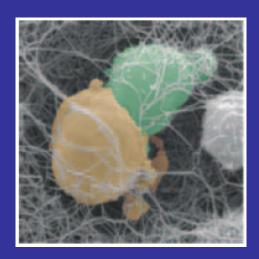




#### Understanding T cell activation is important

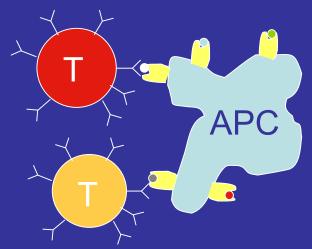
- T cells perform surveillance on antigen-presenting-cells (APCs), looking for signs of invaders
- Upon activation, they can
  - directly kill infected cells
  - activate B cells that secrete antibodies
  - activate macrophage cells





 T cell receptors bind to peptide-MHC complexes (pMHC) on antigenpresenting cells (APC)

#### Each T cell holds ~50,000 identical T cell receptors (TCR).



Specificity: TCR on a particular cell bind well to only a small

class of pMHC

Sensitivity: Some T cells can be at least partially activated

by (very) few pMHC.

Speed: If nothing is recognized, T cell moves on quickly.

TCR signal locally but most expts measure cellular responses:

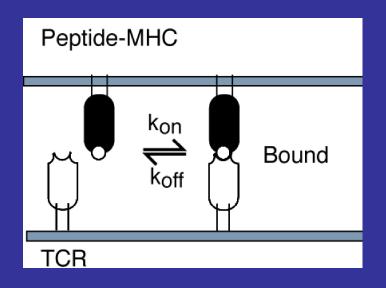
- Cytokine release
- TCR downregulation

# Can we predict the T cell response based on the physical properties of the TCR-pMHC bond?

(the answer has to be YES)

- 1. Introduction
- 2. Measuring bond properties
- 3. Previous models of T cell activation: Serial engagement, kinetic proofreading, et al
- 4. Confinement time model of TCR signaling (2009)
- 5. Theoretical properties of confinement time models

#### Quantifying pMHC-TCR interaction



```
forward rate = k_{on}

reverse rate = k_{off}

half-life t_{1/2} = ln(2) / k_{off}

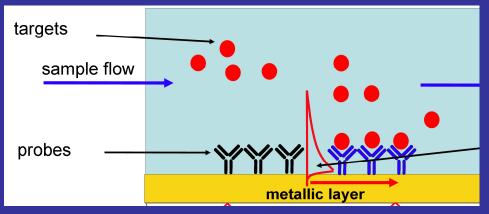
Dissociation constant

= k_{off} / k_{on}

= K_D
```

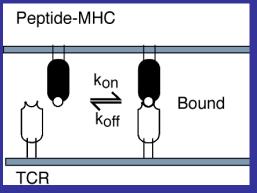
- Measure on and off rates for the reaction using SPR (BIAcore).
- Bonds are found to be weak and transient.

#### Two and three dimensional kinetics



SPR measurements are 3-D:

$$[k_{on}] = M^{-1}s^{-1} = cm^3 s^{-1}$$
  
 $[k_{off}] = s^{-1}$   
 $[K_D] = M = cm^{-3}$ 



At the immune synapse: 2-D constrained kinetics

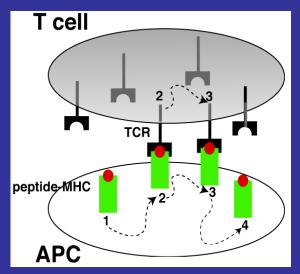
$$[k_{on}] = cm^2 s^{-1}$$
  
 $[k_{off}] = s^{-1}$   
 $[K_D] = cm^{-2}$ 

- Measurements of 2-D kinetics are rare!
- Popular heuristic (probably unreliable!):

$$k_{on}^{2D} = k_{on}^{3D} / L$$
 where L ~ 10nm  
 $k_{off}^{2D} = k_{off}^{3D}$  (receptor scale)

• Also: mechanical forces on bonds at the synapse.

#### Serial Engagement Model



pMHC that bind TCR for a *short* time will bind the most TCR during their time in the contact region.

(high k<sub>off</sub> is good)

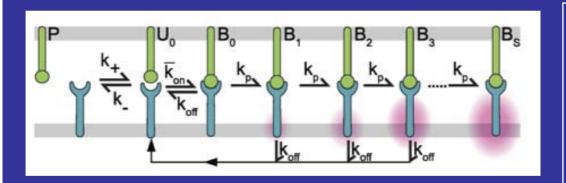
Valitutti et al. (1995) Nature 376:148.

#### Quantity



**Quality** 

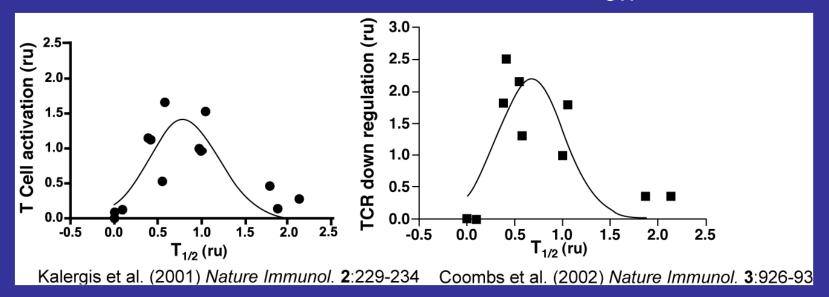
#### Kinetic Proofreading Model

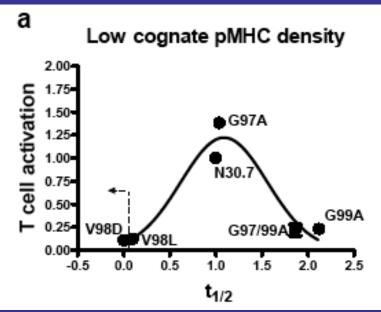


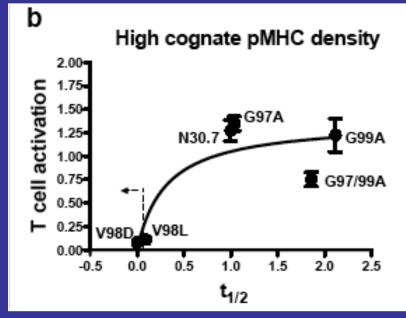
a series of biochemical events leading to full activation must occur before the TCR-pMHC bond breaks (low k<sub>off</sub> is good)

McKeithan 1995, PNAS **92**:5042

#### Experimental support for k<sub>off</sub> models





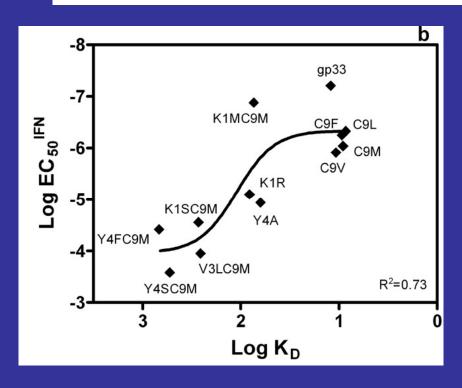


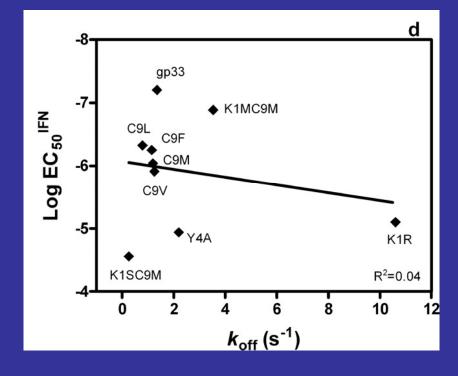
(Gonzalez et al PNAS 2005)

## We focused on the role of the dissociation rate, $k_{\rm off.}$ Other studies underlined the importance of the dissociation constant, $K_{\rm D}$ .

CD8<sup>+</sup> T Cell Activation Is Governed by TCR-Peptide/MHC Affinity, Not Dissociation Rate<sup>1</sup>

Shaomin Tian,\* Robert Maile,\*† Edward J. Collins,\*\* and Jeffrey A. Frelinger<sup>2</sup>\*

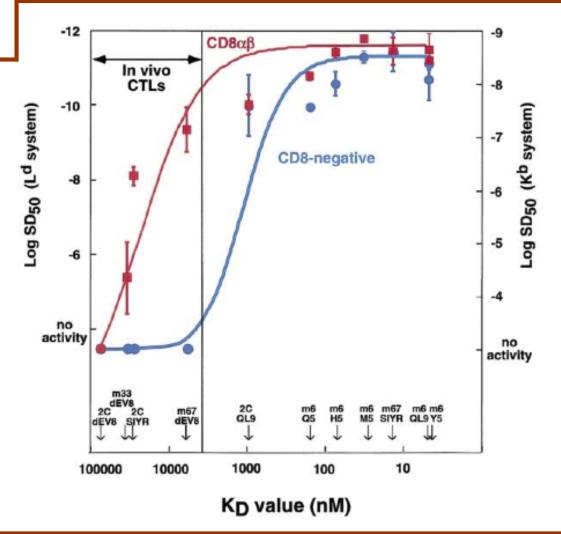




Immunity, Vol. 18, 255-264, February, 2003, Copyright @2003 by Cell Press

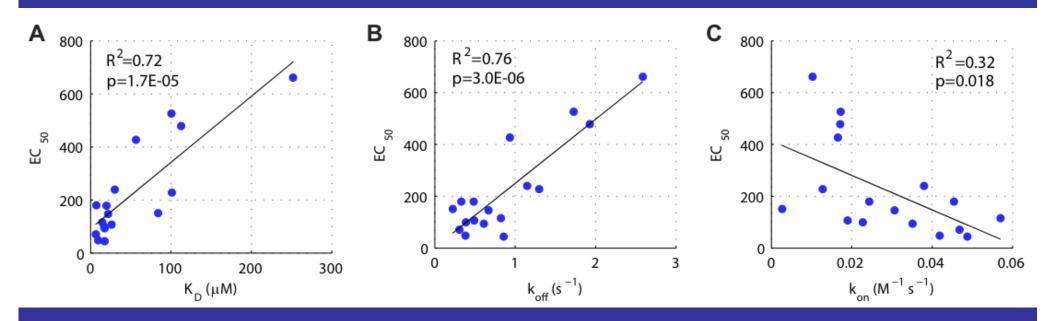
## Quantitative Analysis of the Contribution of TCR/pepMHC Affinity and CD8 to T Cell Activation

Holler and Kranz, Immunity, 2003



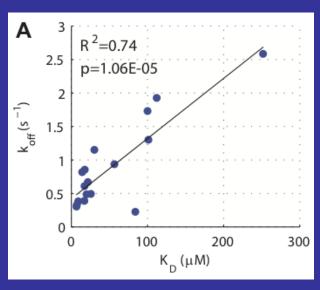
### New experimental work: Aleksic, Dushek et al (Immunity, in press)

- detailed SPR study of 1G4 TCR binding to 17 different altered peptide ligands with wild-type or mutated MHC.
- analyzed the activation of CTL clones by plate-bound pMHC
  - precise control for equal pMHC presentation on plates.

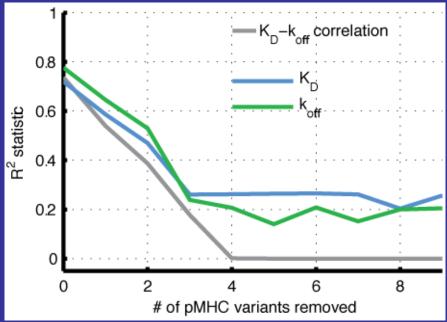


However, there is a confounding correlation between K<sub>D</sub> and k<sub>off</sub>!

#### Subset analysis and bias

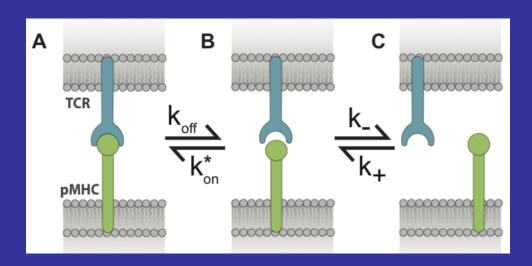


• The correlation between K<sub>D</sub> and k<sub>off</sub> is large.



- We extracted subsets that maximize variation in k<sub>on</sub>.
- K<sub>D</sub> and k<sub>off</sub> fit poorly as subset shrinks.
- Once three pMHC variants are removed, K<sub>D</sub> and k<sub>off</sub> do not correlate significantly with pMHC potency (p>0.05)

#### A new model for surface receptor binding



Membrane diffusion is slow and therefore reactions are likely to be diffusion-limited.

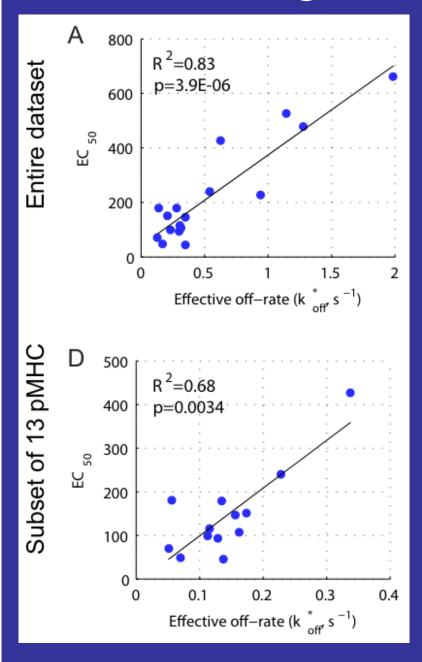
This aspect is missing from some existing models.

$$k_{\text{off}}^* = k_{\text{off}} \left( \frac{k_-}{k_{\text{on}}^* + k_-} \right)$$

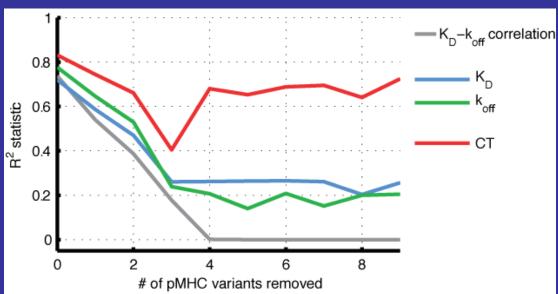
 $\left( \frac{k_-}{k_-^* + k_-} \right)^{\text{K^*}_{\text{on}}} \left( \frac{k_-^*}{k_-^* + k_-} \right)^{\text{K^*}_{\text{on}}} \left( \frac{k_-^*}{k_-^* + k_-^*} \right$ k\*<sub>on</sub> (units of s<sup>-1</sup>) is the intrinsic on-rate; k is the diffusive reverse rate

- if k\*<sub>on</sub> >> k<sub>t</sub> then k\*<sub>off</sub> ~ K<sub>D</sub>
- if  $k^*_{on} << k_{\cdot}$  then  $k^*_{off} \sim k_{off}$

#### Data fitting with the confinement time model



- improved correlation ( $R^2$ =0.83) compared to  $K_D$  and  $k_{off}$ .
- confinement time model R<sup>2</sup> statistic is more robust to removing data points.



• the confinement time model is significant (p<0.05) for all except one data subset.

#### Comparison to previous studies

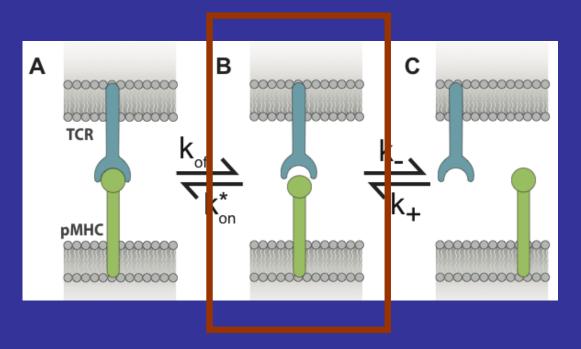
- Many previous studies are ~consistent with the confinement time model.
  - confinement time reduces to k<sub>off</sub> or K<sub>D</sub> under certain conditions.
- When k<sub>on</sub> is large (>10<sup>5</sup> M<sup>-1</sup>s<sup>-1</sup>), pMHC potency correlates well with K<sub>D</sub> (Holler and Kranz, 2003; Tian et al., 2007)
- With smaller  $k_{on}$  (~10<sup>3</sup> M<sup>-1</sup>s<sup>-1</sup>), potency correlates well with  $k_{off}$  (Krogsgaard et al., 2003).
- Our k<sub>on</sub> values were intermediate (~10<sup>4</sup> M<sup>-1</sup>s<sup>-1</sup>).
  - This may explain why the confinement time model provided the best description of our data.

#### Evidence for the confinement time model

- Interactions at cell-cell interfaces last longer than solution measurement predicts. (Grakoui et al., 1999; Tolentino et al., 2008).
  - Lifetime of a CD2-CD58 bond is 100x longer than in solution.
  - However: membrane-tethered interactions could be shorter because of mechanical forces.
  - Direct measurements of 2D TCR/pMHC bond lifetimes will help!
- Increasing pMHC mobility on the cell surface inhibits T cell activation.

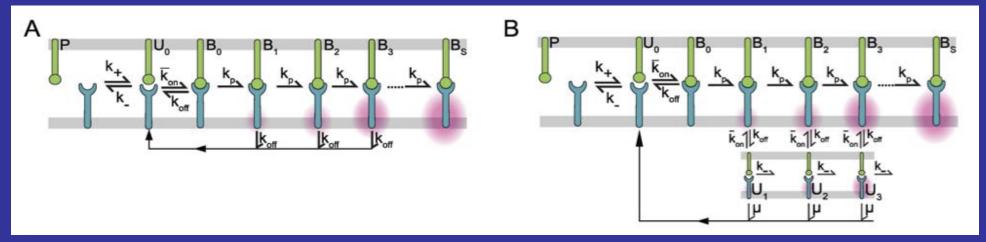
  (Luxembourg et al., 1998; Segura et al., 2008; Wettstein et al., 1991).
  - Confinement time model says that increased mobility decreases rebinding.

#### Confinement time model and signal persistence

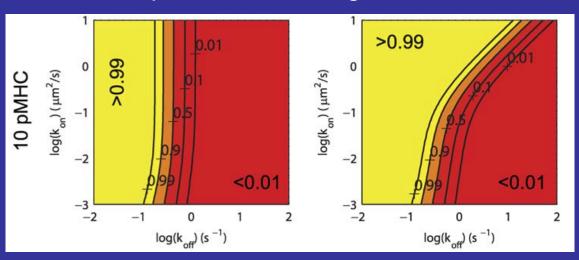


- What is the role of rebinding in antigen discrimination?
- Focus on short-time effects (first 30s of cell interaction).

## A confinement model with signal persistence for **early** TCR signaling



 Modified kinetic proofreading allows signals to persist for a short time after pMHC unbinding.



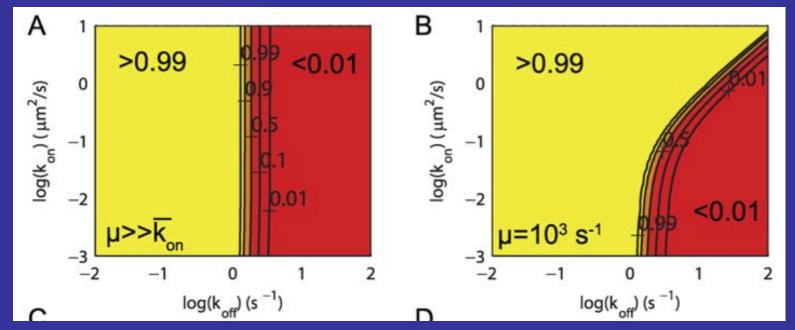
• We find an important role for  $k_{on}$  as well as  $k_{off}$  in signal discrimination.

Probability of one productive signal after 30s interaction.

#### Weak pMHC cannot conspire to signal in this model

(A) no signal persistence

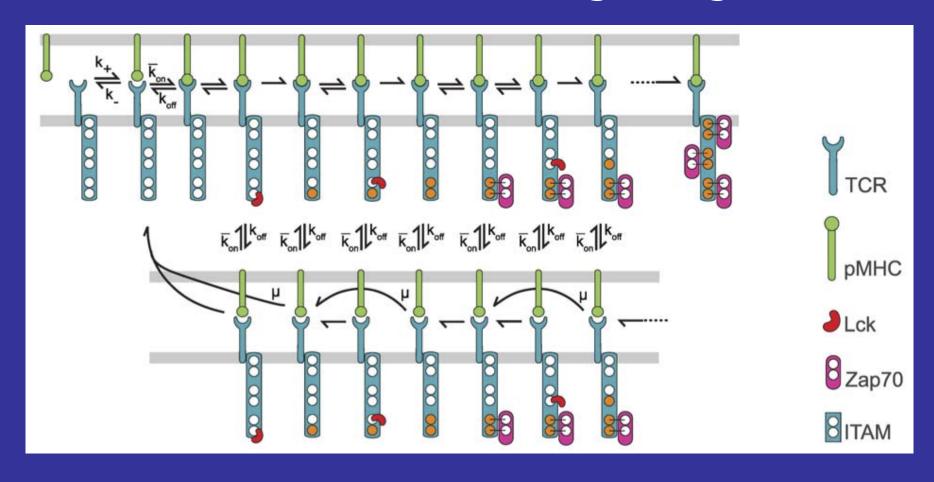
(B) 0.001s persistence



Contours: probability that at least 1 TCR out of 7854 at the contact interface signals within 30s. 39270 identical pMHC are present.

- For short persistence times, weak pMHC (low k<sub>on</sub> / large k<sub>off</sub>) do not signal.
- If signal persistence is long, a sequence of pMHC may activate a few TCR.

## All these findings are recapitulated if we use a more detailed TCR signaling model.



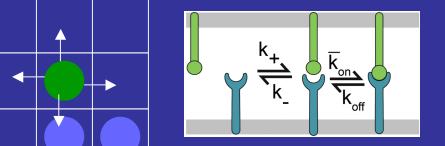
#### Does the ODE model accurately capture the diffusion effect?

1. Formulate a discrete-space continuous-time simulation based on the Gillespie

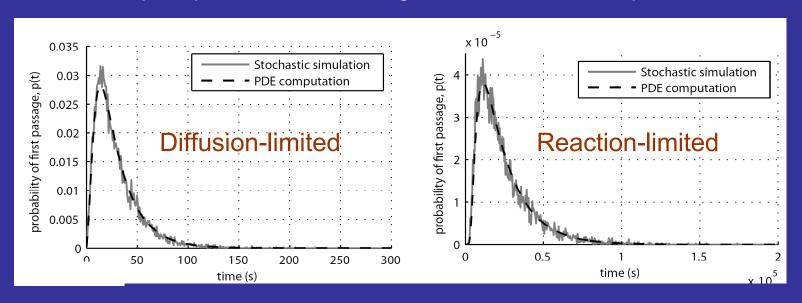
algorithm.

• Single pMHC diffusing on an array of immobile TCR.

Based on the work of Isaacson and Peskin (2006) SIAM Sci. Comp. 28:47-74



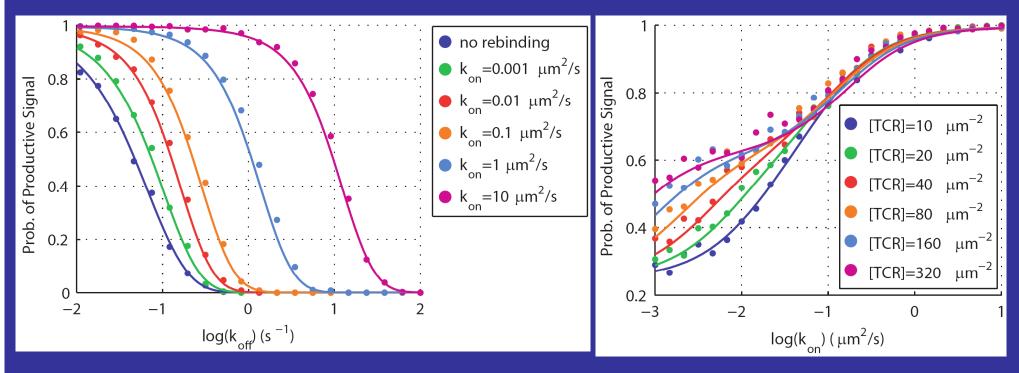
2. This microscopic spatial simulation agrees with macroscopic PDE.



Therefore we have an accurate spatial simulation.

#### ODE model captures the effects of membrane diffusion.

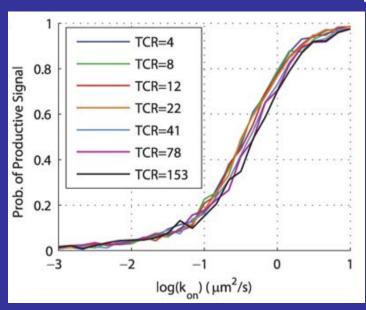
3. Spatial simulations (dots) compare favorably to the ODE formulation (lines).

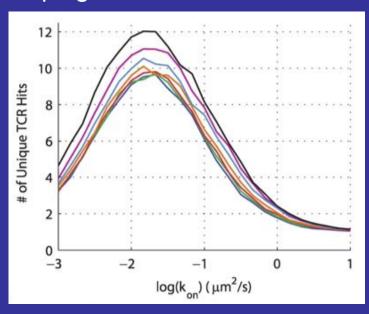


• the ODE formulation accurately captures the effects of membrane diffusion in the parameter regimes we are considering.

#### TCR clustering does not improve signaling

- TCR cluster size has a negligible effect on productive signaling after 30s
- TCR clustering
  - increases the number of unique TCR bound by a pMHC
  - reduces the probability of escaping the cluster





• Underlines the importance of pMHC rebinding to the same TCR versus serial binding of pMHC to different TCR.

[ Spatial lattice Monte Carlo simulation of one pMHC diffusing and binding to TCR starting with one pMHC bound at the center of a TCR cluster (r=100nm). A homogeneous distribution of TCR is assumed outside the cluster. TCR independently perform stochastic kinetic proofreading with signal persistence. The simulation is terminated when t=30s or a productive signal is transduced.]

#### Summary

#### **Experiment:**

- Response of T cells to a panel of 17 pMHC variants.
- Models based on TCR/pMHC confinement time consistently outperform other models.

#### **Modeling:**

- Signal persistence supports rapid and reliable early time antigen discrimination/detection.
  - discrimination on the basis of k<sub>on</sub> and k<sub>off</sub>
  - weak pMHC cannot conspire to signal
- ODE findings are supported by explicit spatial model.
- Findings are robust to using a more detailed TCR signaling model.
- D. Coombs et al. Nature Immunology 3:926 (2002).
- P.A. Gonzalez et al. PNAS 102:4824 (2005).
- O. Dushek, R. Das and D. Coombs PLoS Comp Biol 5:e1000578 (2009).
- M. Aleksic, O. Dushek et al. Immunity, in press (2010).